

Repurposing of Valproic Acid and Simvastatin to potentiate first line chemotherapy regimen

in metastatic pancreatic cancer patients: from preclinical evidence to clinical testing in VESPA trial

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BACKGROUND

Patients with metastatic pancreatic ductal adenocarcinoma (PDAC) have a very poor prognosis, despite all the improvements in cancer therapy [Lambert et al. *Semin Oncol*, 2021], indicating the urgent need for new treatments. Repurposing already-approved non-oncology medications may be a desirable approach in terms of helping to provide efficacious therapeutic alternatives that are easily transferred to early clinical trials.

In tumor models including PDAC, valproic acid (VPA), a generic low-cost anticonvulsant with histone deacetylase (HDAC) inhibitory activity, has been shown to have anticancer characteristics when used alone [Luo D et al. *Carcinogenesis*, 2020] or in combination with gemcitabine [Lin T et al. *JCECR*, 2019].

As we recently shown [Roca MS et al. *JCECR*, 2022], HDAC inhibitors have the ability to sensitize PDAC cells to gemcitabine/abraxane/routa. VPA in combination with conventional chemotherapy is under investigation in different solid tumors, and the results generally support the viability and safety of this strategy [Avallone A et al. *BMC cancer*, 2016; Budillon A et al. *Ann Onc*, 2018].

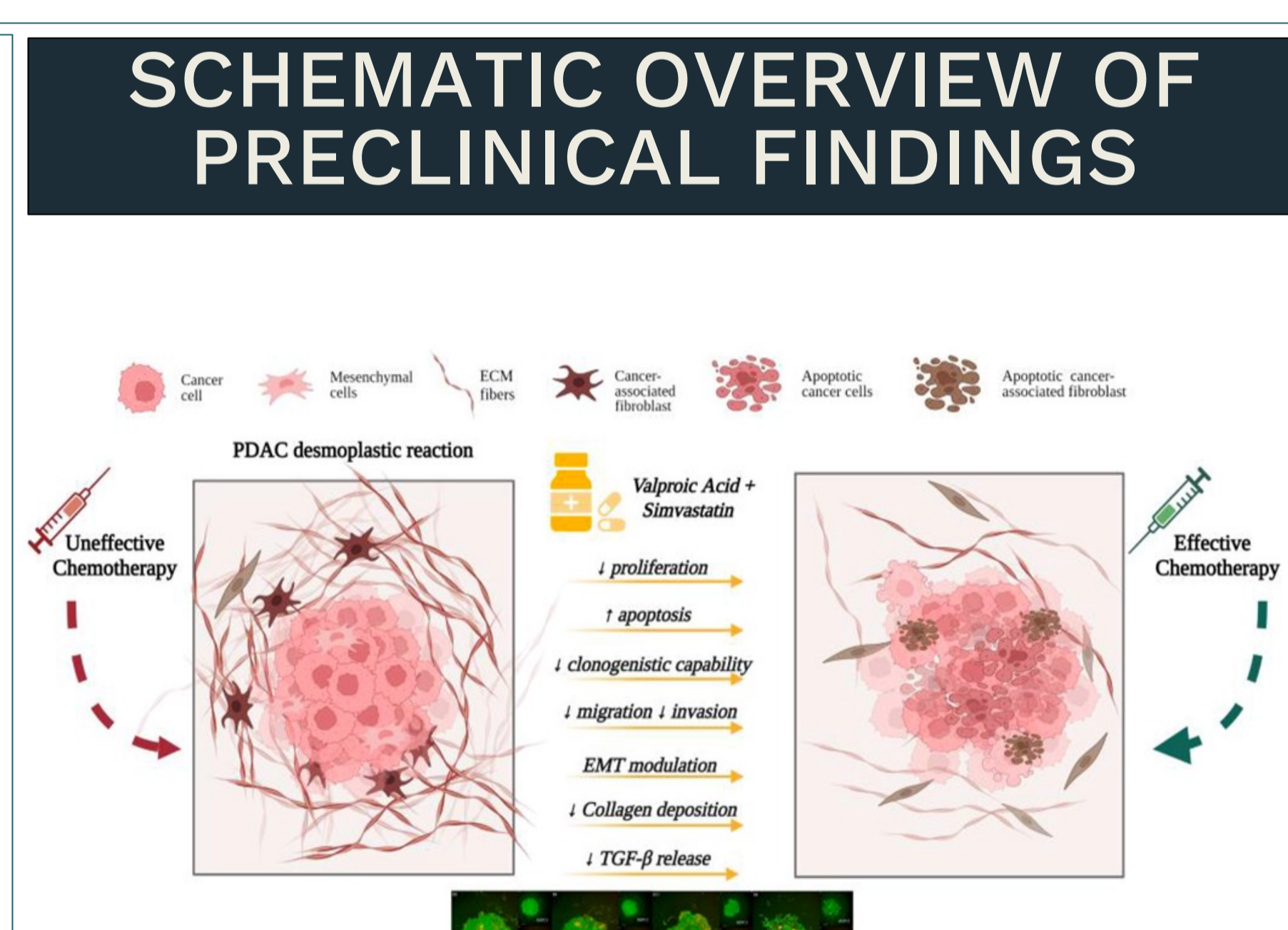
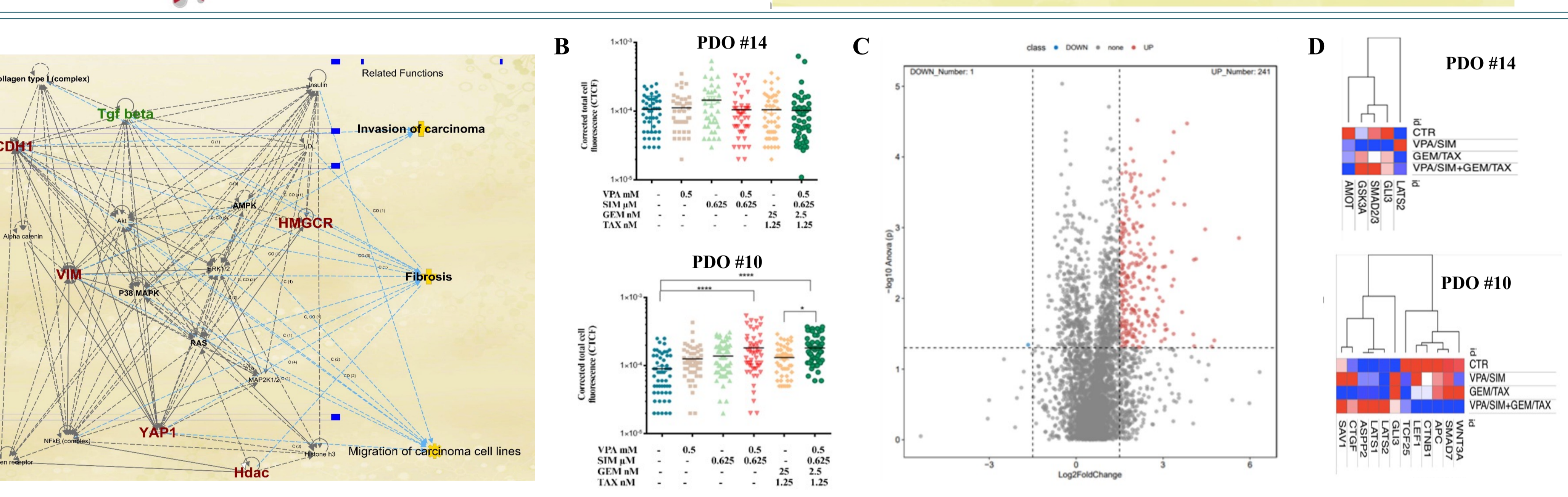
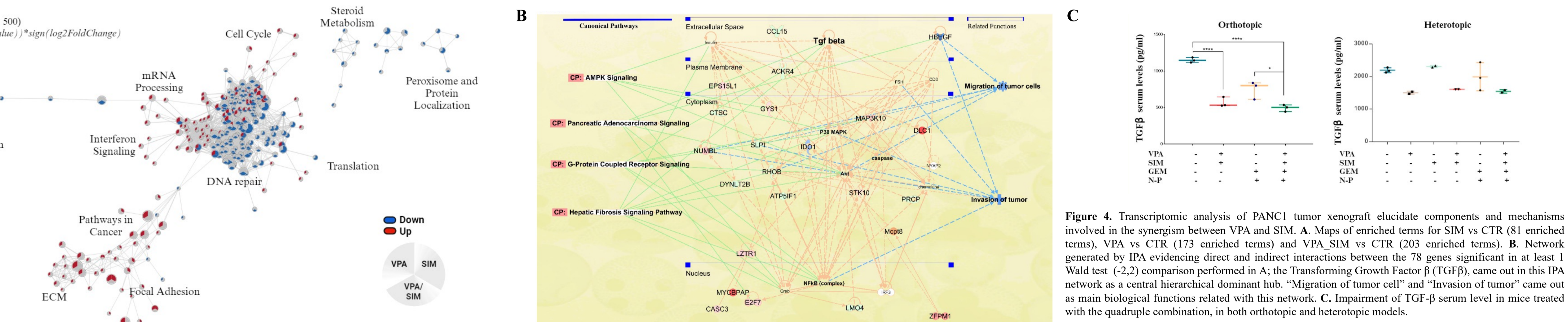
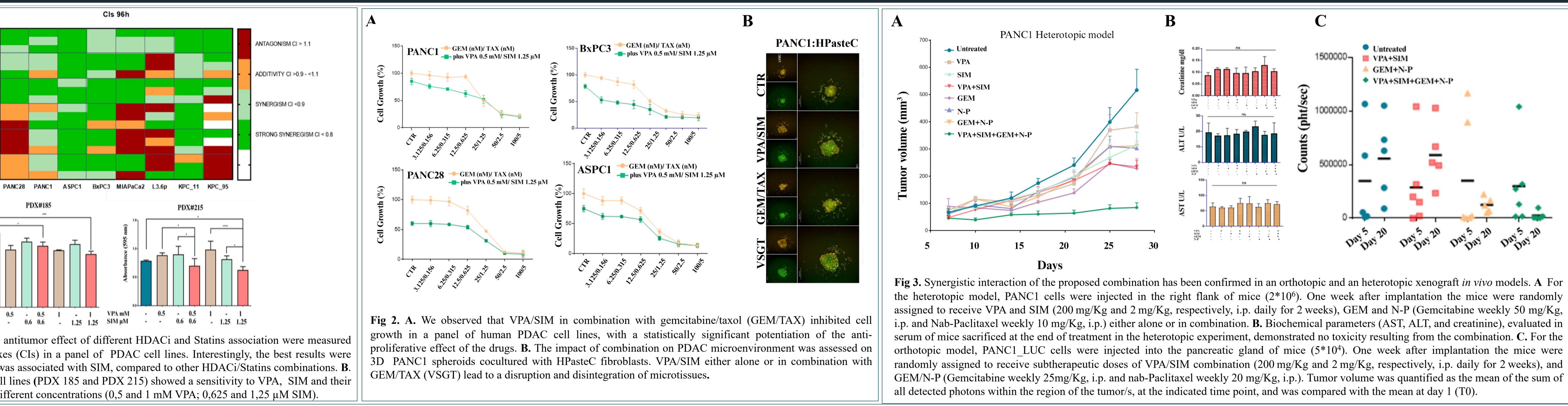
Originally designed to decrease cholesterol by blocking HMG-CoA reductase, statins have also shown a direct antitumor impact when used alone or in combination with chemotherapy and target treatment in a wide range of tumor models, including pancreatic cancer [Gupta V et al. *Cancer Lett* 2018].

We have just demonstrated that VPA and the cholesterol-lowering drug simvastatin have a preclinical synergistic anticancer interaction in metastatic prostate cancer models. Additionally, the combination therapy has the potential to both sensitize prostate cancer cells to docetaxel and reverse docetaxel resistance. This impact has a mechanistic connection to the combined approach's ability to suppress the oncogene YAP and target the cancer stem cells compartment [Iannelli F et al. *JCECR*, 2020].

KEY FINDINGS

- In a panel of human and murine pancreatic ductal adenocarcinoma cells, we demonstrated a strong synergistic antiproliferative and pro-apoptotic effect of valproic acid (VPA) and simvastatin (SIM) combination, either alone or plus chemotherapy. This effect was strengthened by a technical cross-validation carried out in the framework of the EU-funded REMEDI4ALL project.**
- Synergistic antitumor interaction was further observed as impairment of clonogenic capability as well as growth inhibition in 3D models, such as fibroblasts/tumor cell microtissues and patient derived-organoids.**
- The antitumor efficacy has been confirmed in vivo in orthotopic and heterotopic xenograft pancreatic ductal adenocarcinoma models in nude mice.**
- Mechanistically, we also provided evidences that VPA/SIM combination regulate several protumorigenic pathways through TGF-β and YAP signaling modulation, thus potentiating chemotherapy.**
- These findings represent the rationale for the ongoing VESPA trial (EudraCT: 2022-004154-63-NCT: 05821556), a multicentric, patient-centric, open-label, proof-of-concept, "randomized phase 2 study of Valproic acid combinEd with Simvastatin and gemcitabine/nab-paclitaxel-based regimens in untreated metastatic Pancreatic Adenocarcinoma patients," that has already enrolled 32 patients.**
- Overall, we proposed a novel and affordable combination therapy, based on two orally safe and generic drugs, to sensitize a widely employed first-line treatment in poor prognosis mPDAC patients.**

Preclinical evidences of synergistic antitumor effect of VPA/SIM alone or in combination with gemcitabine/taxol



Multicentric, proof-of-concept, open label "Randomized phase 2 study of Valproic acid combinEd with Simvastatin and gemcitabine/nab-paclitaxel-based regimens in untreated mPDAC patients"-The VESPA trial



DESIGN OF THE STUDY

32 Patients enrolled

First-line metastatic PDAC patients

Standard Arm: Gem/Abiraterone or PAXG

Experimental Arm: VPA (1000 mg daily) + Simvastatin (20 mg daily) + Gem/Abiraterone or PAXG

A total of 240 patients, 120 for arm, will be enrolled

SAFETY RUN-IN PHASE

6 patients enrolled in experimental arm receiving AG; 6 patients enrolled in experimental arm receiving PAXG

The study will include an initial 6-patients safety run-in phase in the experimental arm (both for AG and PAXG treatment)

Safety evaluation will be performed by an independent Safety Monitoring Committee.

SCHEMATIC TIMELINE OF STUDY PROCEDURE

Standard ARM: 4 weeks (Gem/Abiraterone or PAXG)

EXPERIMENTAL ARM: 4 weeks (VPA + SIM + Gem/Abiraterone or PAXG)

*AG and PAXG will be administered every 28 days

PATIENT ENGAGEMENT PLAN

Study conduct: Patient recruitment, randomization, follow-up, safety monitoring, quality control, data management, statistical analysis, patient support, patient education, patient feedback.

Analysis: Primary endpoint, secondary endpoints, exploratory endpoints.

Dissemination: Publication, presentation at conferences, patient education, patient support.

WHAT IS PANCREATIC CANCER?

Pancreatic cancer is a disease of the pancreas, a small organ in the abdomen. It starts in the cells that form the pancreas. The cells multiply in an uncontrolled way. This forms a tumor.

Pancreatic Ductal Adenocarcinoma (PDAC) starts in the cells that form these ducts. The cells multiply in an uncontrolled way. This forms a tumor.

Metastatic Pancreatic Ductal Adenocarcinoma (mPDAC) means that your cancer has spread to other parts of your body such as your liver or lungs.

Currently, metastatic PDAC (mPDAC) cancer treatment remains chemotherapy-based but unfortunately, the outcome of this treatment is not good as it is in other types of cancer.

THE VESPA TRIAL AIMS TO:

- Assess if adding two repurposed medications to chemotherapy improves how well this chemotherapy works.
- Slow down the spread of disease.
- Help improve treatment options and outcomes for patients in the future.

STUDY OBJECTIVES

PRIMARY ENDPOINT
PROGRESSION FREE SURVIVAL

SECONDARY ENDPOINTS
OBJECTIVE TUMOR RESPONSE RATE (ORR)
DURATION OF OBJECTIVE RESPONSE (DOR)
DISEASE CONTROL RATE (DCR)
OVERALL SURVIVAL (OS)
OVERALL TOXICITY RATE
QUALITY OF LIFE (QoL)

EXPLORATORY OBJECTIVES
DRUGS PHARMACOKINETICS
METABOLIC PROFILING
CIRCULATING CYTOKINES AND CHEMOKINES
CIRCULATING MICRORNAS EVALUATED AND CPTDNA
PROGNOSTIC AND PREDICTIVE VALUE OF TUMOR DNA MUTATIONAL PROFILING
TRANSCRIPTOMICS PROFILING
PROGNOSTIC AND PREDICTIVE VALUE OF SEVERAL BIOMARKERS EVALUATED BY IMMUNOHISTOCHEMISTRY

CENTERS

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METHODS

Cell Culture The pancreatic cancer cell lines PANC1_ATCC, ASPC1, MiaPaca2, PANC28, B 6pl e COLO_357, and the mouse pancreatic cancer cell lines KPC_11 e KPC_95 were maintained as monolayer cultures and cultured in Dulbecco's modified Eagle's medium (DMEM) containing 4.5 g/L glucose, glutamine, and nonessential amino acids and supplemented with 10% heat-inactivated fetal bovine serum and penicillin (100 IU/mL)-streptomycin (100 µg/mL). BxPC3 cell line were grown in RPMI (Roswell Park Memorial Institute) supplemented with 10% fetal bovine serum (FBS, Cambrex, Belgium) heat-inactivated, 50 units/ml penicillin (Cambrex, Belgium), and 500 µg/ml streptomycin (Cambrex, Belgium). Cell proliferation was measured in cells treated and treated with described drugs as single agent or in combination. Cell proliferation was measured using a spectrophotometric dye incorporation assay (3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT), Sigma-Aldrich). Cell proliferation of PDX-derived cell lines, 185 and 215, was measured using Crystal Violet colorimetric assay and expressed as absorbance at 590 nm. **Microtissues.** PANC1 cells were marked using a fluorescent probe-cell tracker and cultured as microtissues by the ULA System (PerkinElmer). Cancer cells were co-cultured with human pancreatic stellate cells in a ratio of 3:1 as described in literature (Branca V et al. 2020) and untreated or treated with drugs for 96h. 3D microtissues were maintained in the incubator and photographed by Opera Phenix microscope (PerkinElmer) air objective magnification 5X. **In vivo.** All studies have been performed in accordance with "Directive 2010/63/EU on the protection of animals used for scientific purposes" and made effective in Italy by the Legislative Decree DLGS 26/2014. Female athymic nude mice (NCl-nu), which were 6- to 8-weeks old, were purchased from Envigo Laboratories (Huntingdon, UK). The mice were acclimatized in the Animal Care Facility of CROM-Centro Ricerche Oncologiche di Merceglione. To produce pancreatic tumors, PANC1-LUC cells were harvested from subcutaneous cultures and resuspended in PBS solution. RNA-Seq protocol was performed as differential gene expression analysis using Illumina sequencing. Computational analysis including quality control, quantification of gene expression, and differential expression was performed at Mario Negri Institute, Milan. **Patient-Derived Organoids (PDO).** PDOs were generated from tumor surgical procedures of primitive pancreatic cancer specimen. Tumor tissue was processed incubating them in digestion media and plated in matrigel and treated as indicated in the figure. To monitor the sensitivity to different treatments PDO were photographed with an objective magnification 10X and scored by CellEvent Caspase 3/7 Green (Invitrogen, Thermo Fisher Scientific). **Proteomic analysis.** Differentially expressed proteins by Patient-derived organoids at basal level as well as upon treatment was performed by mass spectrometry (LC-MS/MS) and quantified by Progression Q1 for proteomics v. 4.2. Data were filtered using a global FDR < 5% and only proteins with at least one unique identical peptide sequence (p-value < 0.05) were considered identified.

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